

Toxicity Assessment of Structurally Relevant Natural Products from Mexican Plants with Antinociceptive Activity

Karina Martinez-Mayorga,^{1,*} Andrés F. Marmolejo-Valencia,¹ Fernando Cortes-Guzman,¹ Juan Carlos García-Ramos,^{1,§} Eric Iván Sánchez-Flores,¹ Joaquín Barroso-Flores,¹ Jose L. Medina-Franco,² and Baldomero Esquivel-Rodriguez¹

¹ Instituto de Química, Universidad Nacional Autónoma de México, Mexico City, Mexico. C.P. 04510

² Facultad de Química, Departamento de Farmacia, Universidad Nacional Autónoma de México, Mexico City, Mexico.

* Corresponding author: Email: kmtzm@unam.mx, phone: +52-55-5622-4770 (Ext. 4614), fax: +52-55-5616-2217

§ current address: CONACYT-UNAM-Centro de Nanociencias y Nanotecnología. Km 107 Carretera Tijuana-Ensenada. Ensenada, B.C. México. C.P. 22860

Received, December 13th, 2016; Accepted, March 9th, 2017.

Abstract. UNIIQUIM database contains molecules from Mexican plants, one of the richest sources of bioactive molecules in the world. Here, we describe the chemical and toxicological profile of molecules with analgesic activity from UNIIQUIM. Most of the compounds are likely to interact with opioid receptors. The predicted acute toxicity is low and none is predicted mutagenic. Given the structural diversity, and biological and toxicity profiles, these molecules represent a new avenue in the search of molecules with antinociceptive activity.

Keywords: UNIIQUIM; traditional Mexican medicine; analgesics; nociception; toxicity profile.

Resumen. La base de datos UNIIQUIM contiene moléculas de plantas mexicanas, una de las fuentes más ricas de moléculas bioactivas en el mundo. Aquí se describe la química y el perfil toxicológico de moléculas con actividad analgésica obtenidas de UNIIQUIM. La mayoría de los compuestos son propensos a interactuar con los receptores opioides. La toxicidad aguda predicha es baja y ningún compuesto se predice mutagénico. Dada su diversidad estructural y su perfil biológico y toxicológico, son atractivas moléculas líderes con actividad antinociceptiva.

Palabras clave: UNIIQUIM; medicina tradicional Mexicana; analgésicos; nocicepción; perfil toxicológico.

1. Introduction

Mexican flora is among the five richest in the world in terms of diversity and structural complexity of their secondary metabolites including peptides, flavonoids, alkaloids, and terpenes, among others. The first steps towards systematic studies of this chemically and structurally rich content are given by the UNIIQUIM (*Unidad de Informática del Instituto de Química*) group and the CCUD (*Coordinación de Colecciones Universitarias Digitales*) supported by the National Autonomous University of Mexico (UNAM). Natural products play a central role in traditional medicine, nutrition and in drug discovery. Databases of natural products, drugs and nutraceuticals have been created and allow the storage and analysis of chemical information in an efficient and systematic manner. This allows for example assessing diversity and structural novelty, the design of chemical libraries and the qualitative and quantitative studies of structure-activity relationships (SAR, QSAR), the prediction of ADME-Tox properties (absorption, distribution, metabolism, excretion and toxicity). The handling and analysis of chemical databases has been the starting point on drug discovery, the methodologies are general and its use has been expanded to agrochemistry, chemical of flavors, and environmental chemistry, to name few examples.

The use of plants and other natural products as traditional medicine not only conducted to the discovery of opiates as

potent analgesics, but also to compounds with high selectivity towards specific receptors and unusual molecular scaffolds. One of the compounds that had a profound impact in the opioid field is salvinorin A. This compound, obtained and characterized for the first time in 1982 by Dr. Alfredo Ortega in the Institute of Chemistry at UNAM [1], is a psychotropic terpenoid found in the plant *Salvia divinorum*, an indigenous plant from southern Mexico. This molecule is the first known opioid ligand that is not an alkaloid given the fact that it bears no basic nitrogen atoms [2]. This outstanding discovery exemplifies the relevance of natural products and advocates for the exploration and analysis of other compounds of this type.

Better understanding of the different biological mechanisms involved in pain modulation has allowed exploring and designing new molecules with a wide range of affinity and selectivity with the ultimate goal of attenuating undesirable side effects. Molecular modeling studies have contributed in this regard. We are actively working on opioid receptors and its ligands, including those obtained from Mexican plants. For example, we have contributed on the exploration of conformational preferences of opioid peptides into the kappa opioid receptor [3], and the identification of probable binding modes for relevant opioid agonists [4,5]. Molecules with known structural and biological properties are useful as reference. For instance, morphine, herkinorin, DAMGO and naloxone have affinity to the μ -opioid receptor. While morphine is an opiate

benzylisoquinoline alkaloid, mainly extracted from poppy straw of the opium poppy, herkinorin is a semi synthetic analogue of salvinorin A, obtained by deacetylation [6], and DAMGO is a synthetic peptide analogous to enkephalines [7]. In turn, naloxone is an opioid antagonist, it is used to block the effects of opioids, especially in patients who are exposed to overdose [8]. Little to no effect is observed when administered in the absence of opioids. A popular formulation of oxycodone/naloxone is available for controlled release of the former [9]. These molecules represent an opioid-related set of structurally diverse molecules and from diverse sources.

Natural products usually have pharmacological or biological activity. However, some products may be toxic for humans, insects, etc. Thus natural products may show dual roles, depending on the targets for treatment. The toxicity of natural products against cancer cells, viruses, germs, or bacteria has been revised.

The distinct characteristics of the chemical structures of natural products have played a key role in its historic development as lead compounds or sources of active molecules. Key distinct features of natural products are, in general, molecular diversity including coverage in chemical space, structural complexity, and physicochemical properties. These features have been measured and analyzed qualitatively and quantitatively using a broad range of computational approaches.

Recent studies focused on the comparison of natural products with a variety of databases have shown that natural products are distinct in terms of the number of aromatic rings and complex ring systems, the number of chiral centers, the number and proportions of heteroatoms and the degree of saturation [10], have large molecular complexity [11] and adequate drug-like physicochemical properties [12] [13].

Based on the aim to “fail early” in the drug discovery pipeline, it is advantageous to identify and deprioritize molecules

with low probability of success, for instance based on ADME-Tox properties. Scientist and society learned the importance to focus on toxicity as a key aspect on the proposal of new molecules as potential therapeutic or diagnostic agents. An example with devastating consequences is the thalidomide tragedy, reported the mid 60's [14] representing a turning point in toxicity testing. This public health problem prompted to several governments and international regulatory agencies to develop systematic and rigorous toxicity testing protocols on compounds intended for human consumption before going to market. In this sense, the most accepted general toxicity testing guidelines for chemicals are those published by the Organization for Economic Cooperation and Development (OECD). These guidelines are reviewed and updated periodically by expert committees to meet the scientific progress and ever changing assessment practices. Similar recommendations can be found in the fourth revised edition of the Globally Harmonized System of Classification and Labelling of Chemicals (GHS). The most recently revised guidelines for Acute, Sub-chronic and Chronic Oral Toxicity Studies are listed in **Table 1**.

Particularly for pharmaceuticals, the International Conference on Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use (ICH) has developed universally accepted guidelines for acute, sub-chronic, and chronic toxicity testing of pharmaceuticals, which have been accepted by regulatory agencies in all member countries, i.e., US, Europe and Japan (<http://www.ich.org/home.html>). Even though acute toxicity is no longer recommended for development of pharmaceuticals and regardless which of the toxicological test recommendations are being considered, some terms have to be established to determine the toxicity of different substances studied. According to GSH, substances can be allocated on one of five toxicity categories based on acute toxicity numeric cut-off criteria. For oral acute toxicity expressed as

Table 1. Acute, subchronic and chronic oral toxicity study guidelines.

	Guideline	Issue Date	Title	Assay characteristics ^a
Acute	425	03/10/2008	Acute Oral Toxicity Up-and-Down Procedure	Effects produced by a single dose or multiple doses applied in a period of 24 h.
Sub-chronic	407	16/10/2008	Repeated Dose 28-Day Oral Toxicity Study in Rodents	Effects produced by test substance after exposition of 7 days per week for a minimum of 90 consecutive days.
	408	21/09/1998	Repeated Dose 90-Day Oral Toxicity Study in Rodents	
	409	21/09/1998	Repeated Dose 90-Day Oral Toxicity Study in Non-rodents	
Chronic	452	08/09/2009	Chronic Toxicity Studies	Effects produced by an exposition of three months or more
	453	08/09/2009	453 9/8/09 Combined Chronic Toxicity/ Carcinogenicity Studies	

^a Guidance for industry and other stakeholders toxicological principles for the safety assessment of food Ingredients. Redbook 2000. U.S. Department of Health and Human Services. Food and Drug Administration. Center for Food Safety and Applied Nutrition. July 2000; updated July 2007.

medium lethal dose (LD_{50}) is as follows: Category 1: 5 mg/Kg bodyweight, Category 2: 50 mg/Kg bodyweight, Category 3: 300 mg/Kg bodyweight, Category 4: 2000 mg/Kg bodyweight and Category 5: 5000 mg/Kg bodyweight. Medium Lethal Dose (LD_{50}) is the concentration of a substance required to kill half members of a tested population after a specific test duration. Historically obtained by a single dose protocol, nowadays the Up-and Down procedure is employed to decrease the number of animals employed [15]. The toxicity of the compounds can manifest as damage to the genetic material or cellular components, this is associated with the functionality of chromosomes inside cell causing mutations; defined as a permanent change in the amount or structure of the genetic material in a cell. The term mutation applies to both, to heritable genetic changes that may be manifested at the phenotypic level, and to the underlying DNA modification when known, including for example, specific base pair changes and chromosomal translocations. When a compound gives rise to an increased occurrence of mutations in populations of cells and/or organisms, the term mutagenic is used. In both cases, the Ames Assay is one of the most commonly test employed.

During the past 30 years, QSAR methodologies have gradually and greatly evolved [16]. The original use for the quantification of structure-activity relationships widened to environmental toxicology, and QSAR models are now accepted for regulatory purposes [17]. The International Conference on Harmonisation (ICH)-M7 guideline provides a practical framework that is applicable to the identification, categorization, qualification, and control of these mutagenic impurities to limit potential carcinogenic risk [18]. Regarding (Q)SAR models, ICH-M7 states "A computational toxicology assessment should be performed using (Q)SAR methodologies that predict the outcome of a bacterial mutagenicity assay [19]. Two (Q)SAR prediction methodologies that complement each other should be applied. One methodology should be expert rule-based and the second methodology should be statistical-based." (Q)SAR models should follow the general validation principles set forth by the Organization for Economic Co-operation and Development (OECD) [20]. Expert knowledge can be used to review any computer system-based analysis, highlighting the relevance of expert review to provide additional supportive evidence on decision making [17,21,22]. The interpretation of models, in the framework of ICH-M7 is based on structural alerts. Compounds are considered mutagenic or non-mutagenic based on the absence, presence or contradictory structural alerts from two complementary (Q)SAR methodologies (expert rule-based and statistical) allowing to state if further testing is recommended.

To continue our efforts on the study of natural products particularly those with antinociception activity, in this work, we present the chemical and toxicological description of compounds with analgesic properties extracted from Mexican plants. Relevant molecules are included as reference, and the toxicological profile includes the prediction of oral rat IC_{50} values, and mutagenic risk assessment.

2. Results and Discussion

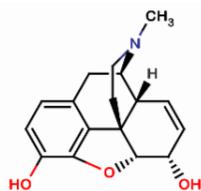
Compounds with analgesic or antinociceptive activity from Mexican plants obtained from UNIQUM are summarized in **Fig. 1**. These molecules include flavonoids, terpenes, alkaloids, etc. For comparison, we included molecules with relevance in analgesia, named: DAMGO, herkinorin, morphine, naloxone, and salvinorin A. These reference molecules, particularly target opioid receptors, which are the main targets for chronic pain, for instance in post-operative and cancer patients.

A great structural diversity is found in compounds with analgesic or nociceptive activity. Literature suggest that the pharmacophore requires the presence of hydrophobic groups, hydrogen bond donors and hydrogen bond acceptors. However, some of the compounds studied in this work do not satisfy this requirement, but show an important analgesic activity with low toxicities.

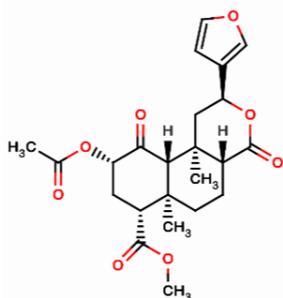
Morphine was the first medicinal alkaloid isolated from any plant and one of the most effective treatment for pain [23]. A study conducted by Dykstra shows that morphine completely blocks acid-induced writhing with a dose of 3.2 (mg/Kg) or 0.011 mmol/Kg [24]. Results for acetaminophen (0.11 mmol, 72.5%), lupenone (0.011 mmol, 50%; 0.023 mmol, 60-65%), epi-friedelanol (0.023 mmol, 45.83%), linarin (0.042 mmol, 38%; 0.337 mmol, 57%), taraxasteryl acetate (0.021%, 77.36%) and caryophyllene oxide (0.056 mmol, 57.87%; 0.113 mmol, 75.19%) were found with the same methodology. None of them show a better activity than morphine, but some show comparable inhibition such as lupenone and taraxasteryl acetate.

Other interesting example is caryophyllene oxide that inhibits the acid-induced writhing in 75%, with a dose ten times higher than that found for morphine, but with a 10-fold decrease in toxicity (caryophyllene oxide LD_{50} = 10.7 mmol/Kg, **Table 3**). Regarding toxicity, all studied compounds showed lower effect than morphine with a decrease of 4 to 10-fold in the LD_{50} values, with exception of acetaminophen, that is practically as toxic as morphine. It is important to note that all studied compounds lack the structural requirements suggested in the literature as key features for their interaction with opioid receptors. However, tests like hot plate or tail-flick suggest that these compounds (linarin, taraxasteryl acetate and caryophyllene oxide) might be targeting opioid receptors. Due to the structural similarity of epi-friedelanol, taraxterol and lupenone with taraxasteryl acetate and similarity found between linarin and acetaminophen is suggested that all compounds shown certain interaction with the opioid receptors, probably with the μ -subtype. Moreover, most of the compounds were predicted as non-mutagenic in both Derek and Sarah, with exception of caryophyllene oxide, for which Sarah gave an out of domain result.

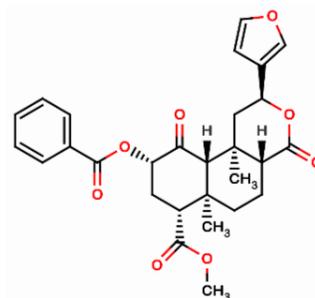
Thus, these compounds isolated from Mexican plants represent a new benchmark in the search for compounds that can interact with opioid receptors, due to the fact that most of them have analgesic or antinociceptive activity with low acute toxicity, most likely by interaction with opioid receptors, without the fulfil of structural requirements previously identified for these kind of compounds.



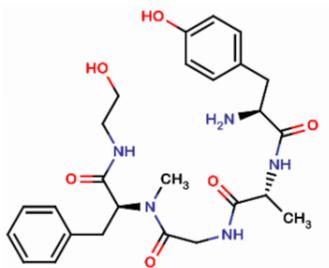
Morphine



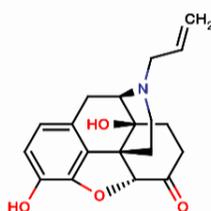
Salvinorin A



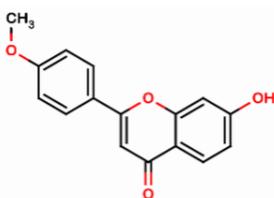
Herkinorin



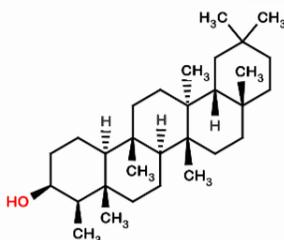
DAMGO



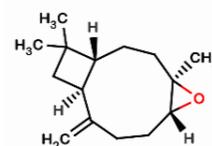
Naloxone



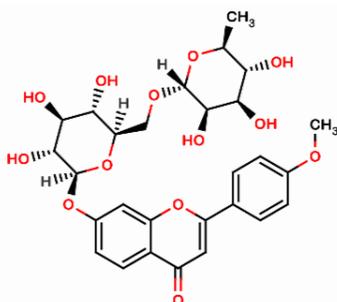
Acacetin



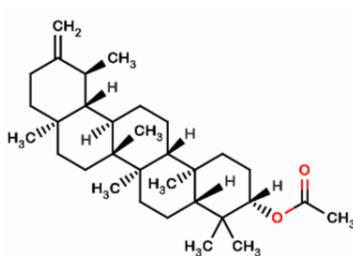
Epi-friedelanol



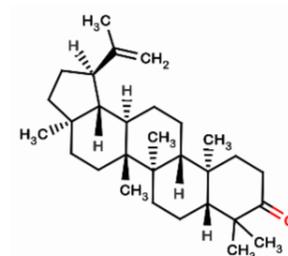
(-)-β-Caryophyllene epoxide



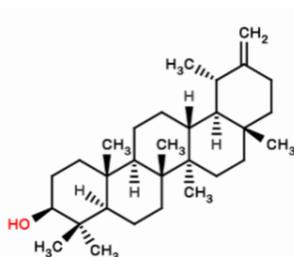
Linarin



Taraxteryl acetate



Lupenone



Taraxasterol anthesterin

Fig. 1. Molecules studied in this work.

The predictions of mutagenicity performed with the software Sarah and Derek from Lhasa Limited are summarized in Table 3. These complementary toxicity predictors are designed to provide the necessary information for decision making under the ICH-M7 guidelines. While Sarah is a statistical-based predictor, Derek uses an expert knowledge-based method. Although both methods have the same endpoint, the way they reach a conclusion is different; therefore they are not redundant but complementary. Results are presented as positive (red) or negative (green) boxes. The associated description substantiates the prediction. In addition, in Sarah a percentage of confidence is provided. Taking together, this information allows to derive a conclusion on the predicted toxicity of each compound.

Based on this data, and according to Lhasa's guidelines, Derek predictions can fall on any of these categories: *Certain*: There is proof that the proposition is true; *probable*: There is at least one strong argument that the proposition is true and there are no arguments against it; *plausible*: The weight of evidence supports the proposition; *equivocal*: There is an equal weight of evidence for and against the proposition; *doubted*: The weight of evidence opposes the proposition; *improbable*: There is at least one strong argument that the proposition is false and there are no arguments that it is true; *impossible*: There is proof that the proposition is false.

If independent methods arrive to similar conclusions to the same endpoint then the accuracy of the results can be increased. For example, when Sarah and Derek are used together the accuracy is as high as 90% [17]. If the independent methods were always in agreement, there will be no need to apply both.

However, when the conclusions are contradicting, the information behind the conclusions need to be analyzed to discriminate why they are contradicting and derive a justified conclusion.

For the reference molecules morphine and naloxone, Sarah predicts they are mutagenic, salvinorin A and herkinorin are equivocal, and DAMGO is predicted inactive with 45% confidence. Among all the other compounds, acacetin is predicted inactive with higher confidence. Caryophyllene oxide is out of domain, and all the other molecules are predicted also inactive with around 25% confidence. In turn, Derek predicts all the compounds inactive, except for caryophyllene oxide that is predicted inactive with unclassified features. This quick but well substantiated overview of the toxicity profile suggests that none of the molecules studied here are predicted to be mutagenic.

Closer look at Derek's prediction for acacetin shows that not only there are not structural alerts for mutagenicity but also there were not unclassified or misclassified features. In turn, Sarah predicts acacetin as negative in a mutagenicity *in vitro* assay with 38% confidence. To arrive to this conclusion, Sarah searches for fragments with known mutagenic risk factor. As an example, **Fig. 2** shows relevant molecules from the training set used within Sarah to come to the conclusion that acacetin will be inactive in the bacterial *in vitro* (Ames) mutagenicity test.

The central analgesic activity of molecules can be studied *in vivo* by means of thermal stimuli with the hot plate test in mice [25]. In turn, peripheral analgesic activity can be evaluated through the acetic acid-induced writhing test *in vivo* [26]. Other ways to study the analgesic or antinociceptive activity are the tail-flick test and formalin-induced paw licking test,

Table 2. Predicted LD₅₀ oral in rat and mutagenicity of compounds with analgesic or antinociceptive activity from mexican plants and reference molecules estimated with "TEST" software.

Compound	-Log10(mol/kg)	Oral rat LD ₅₀ ^a		Mutagenicity ^a	
		mg/kg	mmol/kg		
Morphine	3.08, 2.932 ^b	235.14, 335.28	0.82, 1.17 ^b	0.47, 0 ^b	-, - ^b
Salvinorin A	2.75	772.53	1.78	0.32	-
Herkinorin	2.41	1942.32	3.92	0.31	-
DAMGO	2.36	2234.67	4.23	-0.00	-
Naloxone	2.82	492.65	1.50	0.44	-
Acacetin	3.19	171.37	0.60	0.18	-
Epi-friedelanol	2.29	2204.82	5.14	0.29	-
(-)-β-caryophyllene oxide	1.97	2357.87	10.7	0.09	-
Linarin	2.29	2988.78	5.04	-0.03	-
Taraxasteryl-acetate	2.11	3614.01	7.70	0.12	-
Lupenone	2.45	1522.42	3.58	0.19	-
Taraxasterol	2.29	2202.73	5.16	0.07	-

^a Predicted values.

^b Experimental values. Oral rat LD₅₀: Amount of chemical (mg/kg bodyweight) that causes 50% of rats to die after oral ingestion. Ames mutagenicity: A compound is positive for mutagenicity if it induces revertant colony growth in any strain of *Salmonella typhimurium*.

Table 3. Predicted mutagenicity of compounds with analgesic or noniceptin activity from Mexican plants and reference molecules, estimated with Derek and Sarah Nexus.

	Derek	Sarah (Model - 1.1.19)
Morphine	<p>The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.</p>	<p>The compound is predicted to be positive with 100% confidence for the 'Mutagenicity <i>in vitro</i>' endpoint in the model. This is based on an exact match with a compound found in the training dataset. 3 supporting hypotheses were also found.</p>
Salvinorin A	<p>The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.</p>	<p>The compound is predicted to be equivocal with -- confidence for the 'Mutagenicity <i>in vitro</i>' endpoint in the model. Supporting hypotheses containing similar examples from the training set have been found.</p>
Herkinorin	<p>The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.</p>	<p>The compound is predicted to be equivocal with -- confidence for the 'Mutagenicity <i>in vitro</i>' endpoint in the model. Supporting hypotheses containing similar examples from the training set have been found.</p>
DAMGO	<p>The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.</p>	<p>The compound is predicted to be negative with 45% confidence for the 'Mutagenicity <i>in vitro</i>' endpoint in the model. Supporting hypotheses containing similar examples from the training set have been found.</p>
Naloxone	<p>The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.</p>	<p>The compound is predicted to be positive with 100% confidence for the 'Mutagenicity <i>in vitro</i>' endpoint in the model. This is based on an exact match with a compound found in the training dataset. 4 supporting hypotheses were also found and are displayed for information.</p>
Acacetin	<p>The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.</p>	<p>The compound is predicted to be negative with 38% confidence for the 'Mutagenicity <i>in vitro</i>' endpoint in the model. Supporting hypotheses containing similar examples from the training set have been found. For the hypotheses indicated, the local activity signal generated from the most similar compounds to the query compound contradicts the overall activity signal for the hypothesis.</p>
β -friedelanol	<p>The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.</p>	<p>The compound is predicted to be negative with 25% confidence for the 'Mutagenicity <i>in vitro</i>' endpoint. Supporting hypotheses containing similar examples from the training set have been found.</p>

	Derek	Sarah (Model - 1.1.19)
Caryophyllene oxide	 Inactive ^a The query structure contains features that were not found in the Lhasa Ames test reference set and do not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. It is predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.	 Outside domain At least one fragment derived from the query compound is outside the training dataset domain and an overall prediction is therefore not possible. For those fragments that are in domain, supporting hypothesis is displayed for information.
Linarin	 Inactive The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.	 Negative The compound is predicted to be negative with 24% confidence for the 'Mutagenicity <i>in vitro</i> ' endpoint. Supporting hypotheses containing similar examples from the training set have been found. For the hypotheses indicated, the local activity signal generated from the most similar compounds to the query compound contradicts the overall activity signal for the hypothesis.
Lupenone	 Inactive The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.	 Negative The compound is predicted to be negative with 29% confidence for the 'Mutagenicity <i>in vitro</i> ' endpoint. Supporting hypotheses containing similar examples from the training set have been found.
Taraxasteryl acetate	 Inactive The query structure does not match any structural alerts or examples for (bacterial <i>in vitro</i>) mutagenicity in Derek. Additionally, the query structure does not contain any unclassified or misclassified features and is consequently predicted to be inactive in the bacterial <i>in vitro</i> (Ames) mutagenicity test.	 Negative The compound is predicted to be negative with 24% confidence for the 'Mutagenicity <i>in vitro</i> ' endpoint. Supporting hypotheses containing similar examples from the training set have been found.

^a Contains unclassified features. Specie: bacterium. Endpoint: Mutagenicity *in vitro*.

both in mice [27,28] [29]. Closer look at the biological activity data of the molecules studied in this work shows that all these compounds have been evaluated on either of these pain-related biological endpoints. Table 4 shows *in vivo* biological activity values for each of the molecules studied here. Interestingly, some of these compounds are used in human consumption, for example β -caryophyllene epoxide is a food additive approved by FDA as flavoring substance.

Acacetin is an *O*-methylated flavone isolated from *Robinia pseudoacacia* and *Turnera diffusa* among other plants, it exhibits anti-inflammatory as well as anti-nociceptive properties [30], and is widely investigated as part of the traditional Chinese pharmacopeia [31]. *Robinia* (*R. pseudoacacia*) contains a toxic lectin on its seeds; it has been observed that horses that ingest this plant present anorexia, depression, weakness, and cardiac arrhythmia, among other effects. However, this plant could be investigated for health conditions where these effects are desirable, for instance to investigate its effect on weight management. The flowers however are edible. Acacetin produces a

significant and dose-dependent nociception inhibition of the writhes. Also, acacetin inhibits licking and shaking associated with nociceptive behavior, mainly in the inflammatory phase of the formalin test. At the unique dose of 56.2 mg/kg, shows the maximal efficacy in the writhing and formalin tests, and it was compared to the antinociceptive response produced by the same dosage of tramadol [32].

Lupenone is a derivative of the naturally occurring triterpene lupeol isolated from a variety of plants such as *Acacia visco*, *Abronia villosa* and *Mangiferae Mangifera sp.* As its precursor, lupenone is an antibiotic and anti-inflammatory agent but it has also been found to be cytotoxic against *in vitro* human adenocarcinoma and melanoma models. Interest in developing lupeol-based anti-neoplastic agents, has led to the discovery of highly active derivatives [33]. Intraperitoneal administration of lupenone produces a dose-related and significant inhibition of acetic acid-induced abdominal constriction in mice. Approximately 50% inhibition of nociception is showed with 5 mg/Kg for lupenone; with 10 mg/Kg results in a inhibition of

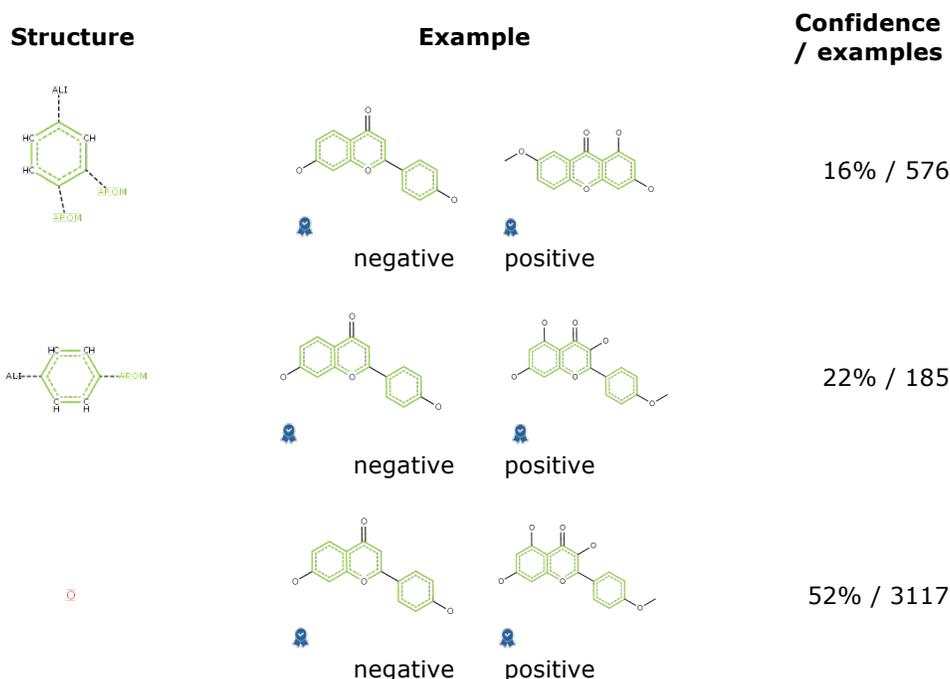


Fig. 2. Hypotheses for acetatin.

nociception ranging from 60 to 65% (Table 4). Pre-treatment of mice with opioid receptor antagonist, completely reverses the antinociceptive effects of lupenone, thus showing the opioid receptor mechanism for nociception. Also, lupenone exhibits 35.6 and 51.2% inhibition of nociception response in the acute and chronic phases, respectively [34].

Epi-friedelanol (also known as *epi*-friedelinol) is a triterpenoid found in a wide variety of plants. It has been reported to exhibit anti-inflammatory [35], antifungal [36] and antileishmanial properties [37]. *Epi*-friedelanol has better inhibition nociception values than diclofenac sodium (10 mg/kg) in the pain's early phase in formalin-induced paw licking test. The inhibition of writhing at 45.83% is considerable for acetic acid-induced writhing test [38].

Linarin is a flavonoid glycoside found in *Valeriana officinalis* which has sedative and sleep-enhancing properties that are potentiated by simultaneous administration of valerenic acid [39], acting as a second order positive modulator of GABA_A receptors. Linarin exerts a significant protective effect from the dose of 50 mg/Kg (45% nociception inhibition) in the acetic acid-induced writhing test, and has a maximum nociception inhibition value at a dose of 200 mg/kg (57%). Linarin (100 mg/kg) significantly increases the reaction time of mice by 55% in the hot plate test. A pretreatment with naloxone antagonized the analgesic effects of the linarin. The inhibitory effect of naloxone, a specific antagonist of morphinic receptors, on the analgesic activity of linarin suggests a morphine-like activity profile [40].

Taraxteryl acetate, a taraxasterol derivative, shows notable antinociceptive activity (Table 4). At 10 mg/kg dose, taraxasteryl acetate exhibits higher activity than acetylsalicylic acid (300 mg/kg) in the tail-flick test, as well as in the acetic acid-induced

writhing tests. Moreover, antinociceptive activity of taraxasteryl acetate (10 mg/kg) is close to that of morphine (10 mg/kg) in the tail-flick test. The highest activity exhibiting taraxasteryl acetate is considered for good antinociceptive properties [41].

(-)- β -caryophyllene epoxide is a sesquiterpene found in most plants from the genus *Cannabis* and is the component which drug-sniffing dogs identify with marijuana [42]. The non-epoxide analogue is widely present in several spices from pepper, cloves, rosemary, basil and lavender; the epoxide is used as food flavoring [43]. Pretreatment with caryophyllene oxide in the hot plate test does not produce any significant changes of paw licking time in the post-treatment early phase. However, in the late phase, the caryophyllene oxide has a dose-dependent and significant increase in licking time treated in mice. The maximum activity is observed with caryophyllene oxide at the 120 min time interval (25 mg/kg bodyweight). Caryophyllene oxide has centrally and peripherally mediated analgesic properties, inhibiting pain in the acetic acid-induced writhing test (Table 4). Further pharmacodynamic investigations are required to understand the analgesic mechanisms [44].

Comparing the reference molecules shown here, the seven molecules under investigation are structurally different, for example, some of them have opioid activity but does not contain all the accepted pharmacophore features for those receptors, such as a positively charge heteroatom and an aromatic moiety. Thus, investigation of binding modes and structural-activity relationship studies are warranted. Given the structural diversity, the nociception activities and the low predicted toxicity of the natural products studied here, they represent a promising hit compounds for further development.

Table 4. Biological activity of compounds with analgesic or noniceptive activity from Mexican plants and reference molecules.

Treatment	Test		Result	Ref
Caryophyllene oxide	acetic acid-induced writhing test	mg/kg wt.	nociception inhibition	[44]
		12.5	57.87%	
		25	75.19%	
	hot plate test	mg/kg wt.	latency to lick the paws after 120 min (sec)	
		control	8.16±1.10	
		12.5	14.5±1.2	
25		17.00±0.96		
Taraxasteryl acetate	acetic acid-induced writhing test	mg/kg wt.	nociception inhibition	[41]
		10	77.36%	
	tail flick test	mg/kg wt.	latency on reaction (sec) 0-30 min	
		control	7.03±0.25	
Lupenone	acetic acid-induced writhing test	mg/kg wt.	nociception inhibition	[34]
		5	50%	
		10	60-65%	
	formalin-induced nociception	mg/kg wt	nociception inhibition	
		10	0-5 min	15-30 min
			35.6	51.2
Linarin	acetic acid-induced writhing test	mg/kg wt.	nociception inhibition	[40]
		25	38%	
		200	57%	
	hot plate test	mg/kg wt.	latency on reaction time (sec)	
		control	11.9±0.9	
Friedelanol	acetic acid-induced abdominal constriction test	mg/kg	nociception inhibition	[38]
		10	45.83%	
	formalin-induced paw licking test	mg/kg	nociception inhibition	
		10	0-5 min	15-30 min
			35.67%	42.94%
Acacetin	acetic acid-induced writhing test	mg/kg	nociception inhibition	[32]
		31.6	72.5	
	formalin-induced paw licking test	mg/kg	nociception inhibition	
		56.2	0-5 min	15-30 min
		50 %		

In conclusion, molecules searched in the UNIIQUIM with nociception activity represent structurally diverse molecules including flavonoids and terpenes. There are indications that lupenone, linarin and taraxteryl acetate exert their nociception effect via opioid receptors. However, for other natural products from Mexican plants with nociception activity the biological pathways involved on this activity need to be explored. Regarding toxicity profile, all the compounds studied do not represent a mutagenicity risk. Some molecules like β -caryophyllene are FDA approved flavor chemicals. The structural diversity of these molecules, their common nociception activity and the predicted safety profile as non-mutagenic agents highlights the importance of these molecules for further studies on the search of analgesic and nociception effects.

3. Methods

Molecules were searched and obtained from the UNIIQUIM database. UNIIQUIM database aims to collect and organize chemical, phytochemical, structural, and biological information of natural products produced for over 75 years in the Institute of Chemistry at the National Autonomous University of Mexico. The database can be searched online at <http://uniquim.iquimica.unam.mx/>. Search of compounds with analgesic or nociceptive activity rendered seven compounds shown in **Fig. 1**.

Prediction of oral LD₅₀ in rat and mutagenicity was performed using TEST (4.1) (Toxicity Estimation Software Tool). In addition, mutagenicity was also predicted with Derek Nexus (v.5.0.2) and Sarah Nexus (v.2.0.1) from Lhasa Limited.

The methodologies implemented in TEST (4.1) are hierarchical clustering, FDA, and nearest neighbors. Details of these methods are described elsewhere [45] and are briefly described here.

Hierarchical Clustering. This method uses Ward's Minimum Variance Clustering Method making a series of n clusters which has only one chemical of n chemicals, so for the training set there will be n clusters, and the variance of test is the sum of the variance of the individual clusters

$$V(l) \equiv \sum_{k=1}^m v(k, l)$$

where $v(k, l)$ is the variance of cluster k at step l . Each of the following steps adds two clusters together to one cluster so the increase of the variance over all clusters is minimized

$$\min V(l+1) \equiv V(l+1) - V(l) = v(k', l+1) - v(k_1, l) - v(k_2, l)$$

where clusters k_1 and k_2 join together at step l to make cluster k' at step $l+1$ the process of combining clusters continues until all of the chemicals are into a single cluster. Then, all clusters are analyzed by a genetic algorithm to determine the optimal descriptor set to obtain the toxicity values of the chemicals within the cluster. The maximum number of descriptors is $n_k/5$.

Prior the genetic algorithm the model are optimized by means of checking for outliers, removing constant descriptors, correlated descriptors, and linearly dependent descriptors, then the predictions are made by "closest cluster from each step" approach in the hierarchical clustering.

FDA. In this method the predictions are made using new clusters constructed containing compounds with at least 75% of similarity coefficient with the test compound. A multiple linear regression model is constructed and the toxicity is predicted, the advantage of this method is that the training set is tailored to fit the test chemical and the disadvantage is that new model has to be generated for each test chemical. If the predictions cannot be made the cluster size is gradually increased from 30 up to 75 chemicals.

Nearest Neighbors. Considered the base line for more complex calculations. In TEST, this method has a fixed similarity coefficient (SC_{min} is set to 0.5). In this method the training cluster for the chemical test is constructed with the three closest chemical relative to the coefficient of similarity of the training set.

Derek and Sarah Nexus, developed by Lhasa Limited, were used to predict bacterial mutagenicity. Settings were left to default values. Derek is the expert toxicity prediction tool. Sarah is a statistical machine-learning methodology for the prediction of bacterial mutagenicity. Molecules are fragmented and reviewed for activity vs. inactivity using self-organizing hierarchical networks, whereby the model arranges relevant fragments. Models arrange relevant fragments into nodes (hypotheses). An overall prediction is obtained based on the prediction and confidence associated with each relevant fragment. Sarah provides a measure of confidence (which is correlated to expected accuracy) for each prediction it makes, for interpretation of a prediction by expert review.

Derek predictions are based on comparison of the information in a chemical structure to the toxicity Derek information or proprietary data in a knowledge base. Derek also highlights fragments of the query compound in order to illustrate the matches to patterns used to hold knowledge contained within Derek; an overall conclusion about the likelihood of toxicity in a structure and detailed reasoning information for the likelihood is so obtained. The prediction is generated by applying expert knowledge rules in toxicology to the data returned from the knowledge base.

4. Acknowledgements

This work was supported by the Institute of Chemistry; and *Facultad de Quimica*, PAIP under Grant number 5000-9163. AFMV and EISF thank to the Consejo Nacional de Ciencia y Tecnología (CONACyT) for providing scholarships. Authors thank ChemAxon, ChemSpider, and Lhasa for providing licenses or online freely available tools. We are indebted to Mizraín Solares, Nikholoz Kobahidze and Noe de la Rosa for providing chemical information. The authors also thank to

Coordinación de Colecciones Universitarias Digitales (CCUD) for financial support.

References

- Ortega A, Blount JF, Manchand PS *J. Chem. Soc., Perkin Trans. 1* **1982**, 2505-2508.
- Ji F, Wang Z, Ma N, Riley J, Armstead WM, Liu R *Brain Res.* **2013**, 1490, 95-100.
- Martinez-Mayorga K, Byler KG, Yongye AB, Giulianotti MA, Dooley CT, Houghten RA *Euro. J. Med. Chem.* **2013**, 66, 114-121.
- Yongye AB, Appel JR, Giulianotti MA, Dooley CT, Medina-Franco JL, Nefzi A, Houghten RA, Martinez-Mayorga K *Biorg. Med. Chem.* **2009**, 17, 5583-5597.
- Marmolejo-Valencia AF, Martinez-Mayorga K *J. Comput. Aided Mol. Des.* **2017**, 31, 467-482.
- Xu H, Partilla JS, Wang X, Rutherford JM, Tidgewell K, Prisinzano TE, Bohn LM, Rothman RB *Synapse* **2007**, 61, 166-175.
- Root J, Wakenhut AMLC, Siggins H, Katugampola S, Napier C *Naumyn-Schmiedeberg's Arch. Pharmacol.* **2012**, 385, 1049-1052.
- Kim HK, Nelson LS *Expert Opin. Drug Saf.* **2015**, 14, 1137-1146.
- Frampton JE *CNS Drugs* **2015**, 29, 511-518.
- Feher M, Schmidt JM *J. Chem. Inf. Comput. Sci.* **2003**, 43, 218-227.
- López-Vallejo F, Giulianotti MA, Houghten RA, Medina-Franco JL *Drug Discov. Today* **2012**, 17, 718-726.
- Medina-Franco JL, Martinez-Mayorga K, Meurice N *Expert Opinion on Drug Discovery* **2014**, 9, 151-165.
- González-Medina M, Prieto-Martínez FD, Naveja JJ, Méndez-Lucio O, El-Elimat T, Pearce CJ, Oberlies NH, Figueroa M, Medina-Franco JL *Future Med. Chem.* **2016**, 8, 1399-1412 DOI: 10.4155/fmc-2016-0079.
- Kim JH, Scialli AR *Toxicol. Sci.* **2011**, 122, 1-6.
- Auletta CS (2014). *General Toxicology in Handbook of Toxicology* Boca Raton, FL: CRC Press.
- Walker JD *QSAR & Combinatorial Science* **2003**, 22, 415-421.
- Barber C, Amberg A, Custer L, Dobo KL, Glowienke S, Van Gompel J, Gutsell S, Harvey J, Honma M, Kenyon MO, Kruhlak N, Muster W, Stavitskaya L, Teasdale A, Vessey J, Wichard J *Regul. Toxicol. Pharmacol.* **2015**, 73, 367-377.
- ICH-M7 International Conference on Harmonisation.
- ICH-M7 (2015). M7 Assessment and Control of DNA Reactive (Mutagenic) Impurities in Pharmaceuticals to Limit Potential Carcinogenic Risk Guidance for Industry.
- Retrieved April 20, 2015, from <http://www.oecd.org/chemicalsafety/risk-assessment/validationofqsarmodels.htm> **2004**.
- Dobo KL, Greene N, Fred C, Glowienke S, Harvey JS, Hasselgren C, Jolly R, Kenyon MO, Munzner JB, Muster W, Neft R, Vijayaraj Reddy M, White AT, Weiner S *Regul. Toxicol. Pharmacol.* **2012**, 62, 449-455.
- Rusyn I, Daston GP *Environ. Health Perspect.* **2010**, 118, 1047-1050.
- Hayes AN, Gilbert SG (2009). Historical milestones and discoveries that shaped the toxicology sciences. In *Molecular, Clinical and Environmental Toxicology*, Vol. 1, 20 (Ed Lunch A.). Basel, Switzerland: Birkhäuser Verlag.
- Miller LL, Picker MJ, Schmidt KT, Dykstra LA *Psychopharmacology* **2011**, 215, 455-465.
- Eddy NB, Leimbach D *Journal of Pharmacology and Experimental Therapeutics* **1953**, 107, 385-393.
- Koster R, Anderson M, De Beer EJ *Fedn. Proc.* **1959**, 18, 412-418.
- D'Amour FE, Smith DL *J. Pharmacol. Exp. Ther.* **1941**, 72, 74-79.
- Davis PHPH, Davis P (1975). *Flora of Turkey and the East Aegean Islands. Vol.5*. Edinburgh University Press.
- Hunnskaar S, Hole K *Pain* **1987**, 30, 103-114.
- Ana G, Eduarda F, Jose LFCL, Lurdes M, Corvo ML *Curr. Med. Chem.* **2008**, 15, 1586-1605.
- Yuan-yuan Z, Bo-hua Z, Wei-guo S *Chinese Journal of New Drugs* **2014**, 1053-1056.
- Carballo-Villalobos AI, González-Trujano ME, López-Muñoz FJ *Eur. J. Pain* **2014**, 18, 396-405.
- Bhui K, Srivastava AK, Shukla Y (2010). Cytotoxic action of natural pentacyclic triterpenes on cancer cells: lupane-type compounds. In *Pentacyclic Triterpenes as Promising Agents in Cancer* (Ed Salvador J. A. R.). Azinhaga de Santa Comba, Portugal: Nova Science Publishers.
- Talla E, Dabole B, Taiwe GS, Ngo Bum E, Mbafor JT, Atchade ADT, Malik R, Zulfiqar A, Sidiki N, Nguimbou RM, Choudhary MI *Pharmacologia* **2013**, 4, 218-227.
- Ramesh N, Viswanathan MB, Saraswathy A, Balakrishna K, Brindha P, Lakshmanaperumalsamy P *J. Ethnopharmacol.* **2002**, 79, 129-132.
- Viswanathan MB, Ramesh N, Ahilan A, Lakshmanaperumalsam Y *P Med. Chem. Res.* **2004**, 13, 361-368.
- Yamamoto ES, Campos BLS, Jesus JA, Laurenti MD, Ribeiro SP, Kallás EG, Rafael-Fernandes M, Santos-Gomes G, Silva MS, Sessa DP, Lago JHG, Levy D, Passero LFD *PLoS ONE* **2015**, 10, e0144946.
- Ghosh S, Chattopadhyay D, Mandal A, Kaity S, Samanta A *Med. Chem. Res.* **2013**, 22, 4347-4359.
- Fernández S, Wasowski C, Paladini AC, Marder M *Pharmacol. Biochem. Be* **2004**, 77, 399-404.
- Martínez-Vázquez M, Ramírez-Apan T, Aguilar H, Bye R *Planta Med.* **1993**, 62, 137-140.
- Bahadır Ö, Çitoğlu GS, Šmejkal K, Dall'Acqua S, Özbek H, Cvacka J, Zemlicka M *J. Ethnopharmacol.* **2010**, 131, 83-87.
- Ross SA, ElSohly MA *J. Nat. Prod.* **1996**, 59, 49-51.
- Efsa Panel on Food Contact Materials EF, Processing A *EFSA Journal* **2014**, 12, n/a-n/a.
- Chavan MJ, Wakte PS, Shinde DB *Phytomedicine* **2010**, 17, 149-151.
- Martin TM, Harten P, Venkatapathy R, Das S, Young DM *Toxicol. Mech. Methods* **2008**, 18, 251-266.